

Studies on the Phyllosoma Larva of the Indian Rock Lobster, *Panulirus Homarus* Linnaeus, 1758

S. Lazarus, J. C. Nisha, R. Thangaraja

Institute for Environmental Research and Social Education, Nesamony Nagar, Nagercoil, Tamil Nadu, India

ABSTRACT

Attempts were made to develop a technique to rear the phyllosoma larvae of *Panulirus homarus*. The biological characters like fecundity, hatching percentage, larval morphological changes, feed inputs and moulting frequency till the fourth moult were studied. Morphometric and meristic characters of the larvae were also studied till the 42nd day. The larval output was directly proportional to the size of the gravid brood stock. Relationship between the duration of culture (X) and length of the larvae (Y) were shown by the relationships $Y \text{ intercept} = -0.5780 \pm 0.1074$ and $X \text{ intercept} = 0.7283$ ($r^2 = 0.8519$). There was significant ($p < 0.0001$) positive relationship between total length (TL) and carapace width (CW) of phyllosoma larvae.

KEYWORDS: *Phyllosoma; spawners; morphometric; meristic; fecundity; moulting*

How to cite this paper: S. Lazarus | J. C. Nisha | R. Thangaraja "Studies on the Phyllosoma Larva of the Indian Rock Lobster, *Panulirus Homarus* Linnaeus, 1758" Published in International Journal of Trend in Scientific Research and Development (ijtsrd), ISSN: 2456-6470, Volume-4 | Issue-4, June 2020, pp.1740-1744, URL: www.ijtsrd.com/papers/ijtsrd31676.pdf



Copyright © 2020 by author(s) and International Journal of Trend in Scientific Research and Development Journal. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0) (<http://creativecommons.org/licenses/by/4.0>)



INTRODUCTION

Aquaculture practice is gaining wide spread importance all over the world as a source of aquatic animal protein. Lobsters, crabs and scallops continue to attract much research attention today because of their demand. The success of aquaculture depends greatly on the development of larval rearing technique. For hatchery production of any commercially important larvae, the stage-wise identification during metamorphosis is very important. The larvae can be fed with a particular feed only after identifying the larval stages (Lazarus *et al.*, 2000). The commercial, recreational and traditional values of spiny lobsters in many countries have led to considerable scientific study, particularly in *Panulirus spp.* But larval development of *Panulirus homarus* is still poorly understood.

The spiny lobster, *Panulirus homarus*, is having a very high economic value and demand. But its production through hatcheries is not possible yet because of the poor understanding of the larval development and behaviour. The larvae exhibit a long pelagic and planktonic phase, which makes it difficult to maintain in the hatchery, under controlled conditions.

The first success in rearing a palinurid lobster from egg to peurulus stage was achieved by Kittaka (1988), Kittaka and Ikegami, (1988) and Kittaka *et al.* (1988). Malachite green exposed phyllosoma larvae were examined through six developmental stages by nine moults. Radhakrishnan and

Vijayakumaran (1995). Maharajan *et al.* (2012) studied the fecundity and viability of eggs of spiny lobsters, *Panulirus homarus*. The survival and growth of phyllosoma larvae of the spiny lobster, *Panulirus homarus* under different salinity regimes were studied by Dilip *et al.*, (2010). Captive breeding of the spiny lobster, *Panulirus homarus* was studied by Vijayakumaran *et al.*, (2005). In the present work, an attempt was made to study the fecundity, hatching percentage, larval morphological changes and moulting frequency of the larvae till the fourth moult were made.

MATERIAL AND METHODS

The exogenous spawners of *Panulirus homarus* were collected from a landing centre in Muttom, south west coast of India. Three stages of spawners based on the colour of the egg mass such as orange, reddish orange and pale brown were selected as source of spawners for the present study. The disease free and healthy spawners having dark brown eggs were used for spawning. The collected spawners were brought to the hatchery by using a special transportation device fabricated by Babu and Marian (1998) and acclimatized to the hatchery conditions. After acclimatization, the brooders were dipped in fresh water for 30 seconds to eliminate external fungi and bacteria. After that they were treated with 10 ppm formalin for 2 min. The treated brooders were again washed in fresh sea water to remove the formalin effect found on the exogenous eggs following Babu *et al.*, (2001a).

The brooders of first two maturation stages, with orange and red coloured eggs, were not stocked in the hatching tank. They were maintained in the maturation tank and fed with clams and muscles and water exchange till the egg reached the third stage (i.e.,) the brown coloured egg stage. The 3rd stage exogenous animal with dark brown eggs obtained both from natural source and maturation tanks were transferred in to the hatching tank for the release of young ones from the eggs. In the maturation tank, the brooders were fed with squid and mussel meat at a rate of 20% body weight and 50% water exchange was carried out daily. The stress during this stage was minimized in order to prevent the premature shedding of eggs.

The hatching was confirmed in the tanks by the vigorous pleopod movements of the mother lobster and following larval release in the hatching tank. The hatched larvae were separated intermittently from the spawning tank and stocked in larval rearing tanks of 200 L capacity at the rate of 10 nos./L stocking density, where the larvae were fed with *Nannochloropsis*, an ultra plankton, at the rate of 1 lakh cells/ml concentration. From the second day onwards, the larvae were fed with newly hatched *Artemia* nauplii and micro algae. The physical parameters maintained during the culture were temperature $30 \pm 2^\circ \text{C}$, salinity $35^\circ/\infty$, dissolved oxygen 5 mg/l and pH 8.0 ± 2 (Lazarus *et al.*, 2000).

Following the method of Babu *et al.* (2001a), cradle aeration system was improvised for a homogenized and adequate air supply to the rearing system. About 50-90% of sea water exchange was carried out daily. The morphological changes observed in antennules, antenna, head, eye, eyestalk, I maxilliped, II maxilliped, III maxilliped, I, II, III and IV pereopod, setae, spines and cephalothorax were observed using light microscope and all the morphological variations were drawn using camera lucida. The fecundity, larval moult, growth rate and survival rates were also recorded.

The fecundity was recorded in relation with spawner size. The exogenous spawners of a visually acceptable size were allowed to release their eggs as hatched out larvae. After the release of larvae, both released larvae and unhatched eggs were counted. From the counted value, the total fecundity were calculated. Percentage of survival was calculated on the basis of daily mortality. The growth rate was determined microscopically by measuring the total length (TL) and carapace width (CW) of larvae during every moult.

Statistical Analysis:

Coefficient of regression analysis was performed for total length (CL) and carapace width (CW). Data analysis were based on mean \pm SEM. Turkey's Multiple Comparison Test was performed for moulting frequency and fecundity with 95% confidence level. This statistical analysis was performed by using the software Graph Pad Version 4.0.

RESULTS

Metamorphosis

Before 1st moult

In the newly hatched larvae, the head was cone-shaped. The antennules and antennae were uniramous. The eyes were without eyestalk. Mandibles slightly developed. First and second maxillae were seen as rudiments. Second maxilliped was uniramous with 4 segments. First pereopods were biramous and segmented. Endopod 3 segmented, ends with

long narrow spine, surrounded by a group of 7 small spines. The terminal segment was spiny and the 2nd pereopod rudimentary (Fig. 1).

After 1st moult

Head was slightly narrower at the anterior region and broader at the posterior region. Antennule uniramous, spiny and unsegmented with 3 aesthetases and 2 setae in the terminal region. Antennae uniramous, unsegmented with 2 pairs of setae. Distinct eyestalk was observed. Mandibles slightly developed. 1st and 2nd maxilla were seen rudimentary. 2nd maxilliped uniramous, four segmented. Last segment had 2 setae and third segment had 6 setae. 1st pereopod biramous, endopod 3 segmented, ends in a long narrow spine, endopod 7 segmented with 6 pairs of plumose setae and the 4th pereopod seen as rudiment (Fig.2).

After 2nd moult

Head, narrow in the anterior region and broader at the posterior region. Antennae and antennules uniramous. Distinct eye stalk. 1st and 2nd maxilla rudiments, mandibles slightly developed. 1st maxilliped uniramous, 2 segmented. Second maxilliped uniramous, 5 segmented and segments had 3 setae. Third segment had 6 setae. First pereopod biramous, exopod 8 segmented with 7 pairs of plumose setae. Towards the end of the stage, 9 segment and 8 pairs of plumose setae were seen. Fourth pereopod was rudimentary (Fig. 3).

After 3rd moult

Head, narrower in the anterior region and broader at the posterior region. Antennae and antennules uniramous. Distinct and elongated eye stalk. First and second maxilla and mandibles slightly developed. Second maxilliped uniramous, 5 segmented, last segment had 3 setae and third segment had 6 setae. First pereopod biramous, exposed, 10 segmented with 9 pairs of plumose setae. Fourth pereopod slightly developed (Fig.4).

After 4th moult

There was no change in the shape of cephalic shield, antennae and antennules, distinct and elongated eye stalk. 1st and 2nd maxillae and mandibles slightly developed. First maxillipede 3 segmented, second maxillipede uniramous and 5 segmented, last segment had 3 setae. Third maxillipede bigamous with exopod setae. First and second pereopods biramous, 7 segmented with 9 pairs of plumose setae in the exopod. Endopod ends with long narrow spine surrounded by a group of small spines. The terminal segment was spiny, and the third pereopod was biramous with five pairs of exopod setae. Fourth pereopod slightly developed and bi-segmented (Fig. 5).

Growth rate

The growth parameter of the relationship of total length (TL) and carapace width (CW) of phyllosoma larvae is shown in the Fig. 6. Measurement of total length and carapace width of the larvae was carried out up to 42 days. The newly hatched larvae had a total length and carapace width of 1.3280 ± 0.0360 mm and 0.6241 ± 0.0561 mm respectively. On the 11th day, the larval length and width observed were 1.6144 ± 0.1290 mm and 0.7960 ± 0.063 mm respectively. On 21st day the measured length and width were 1.7004 ± 0.0993 mm and 0.8266 ± 0.0450 mm respectively. On the 31st day the standard length and width of the larvae observed were 2.283

± 0.0116 mm and 1.0346 ± 0.0565 mm respectively and on the 41st day the standard total length and carapace width of the larvae observed were 2.894 ± 0.6210 mm and 2.017 ± 0.1432 mm respectively (Fig. 6). The slope of the regression data showed positive and highly significant ($p < 0.0001$) relationship between total length (TL) and carapace width (CW) of phyllosoma larvae.

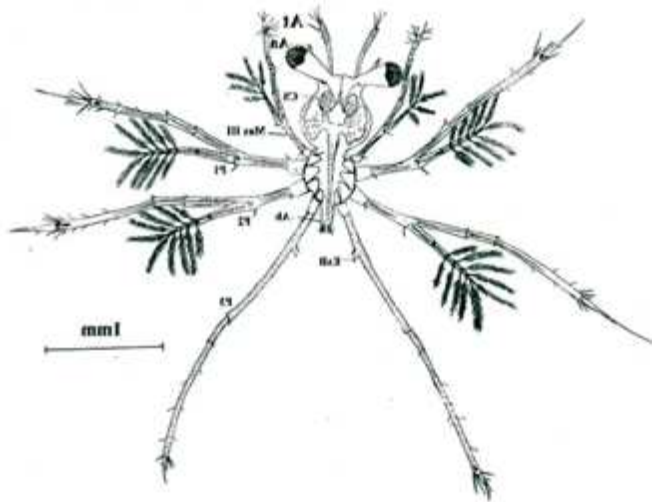


Fig.1. Newly hatched Phyllosoma larva

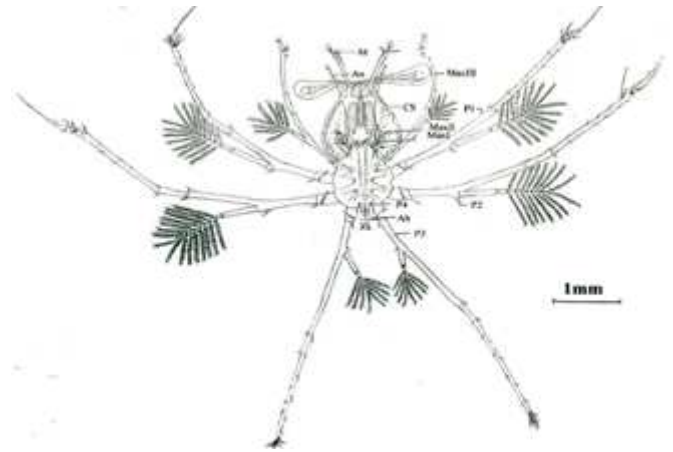


Fig.4. Phyllosoma larva after 3rd moult

At – Antenna, An – Antennule, Max I – Maxillipede I, Max II – Maxillipede II, Max III – Maxillipede III, P1 – Pereiopod I, P2 – Pereiopod II, P3 – Pereiopod III, P4 – Pereiopod IV, Ab – Abdomen, CS – Cephalic shield

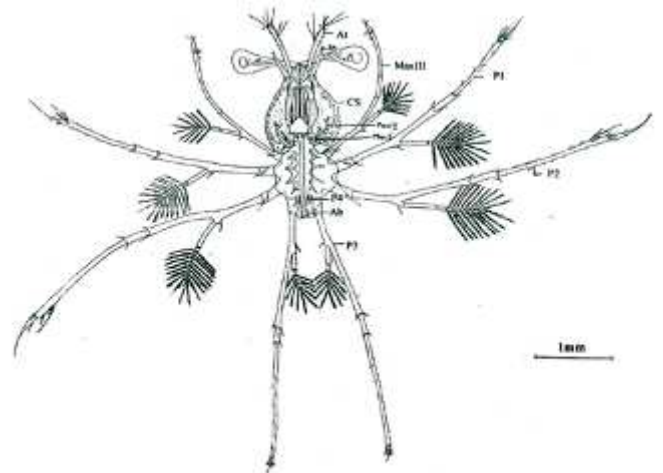


Fig.5. Phyllosoma larva after 4th moult

At – Antenna, An – Antennule, Max I – Maxillipede I, Max II – Maxillipede II, Max III – Maxillipede III, P1 – Pereiopod I, P2 – Pereiopod II, P3 – Pereiopod III, P4 – Pereiopod IV, Ab – Abdomen, CS – Cephalic shield

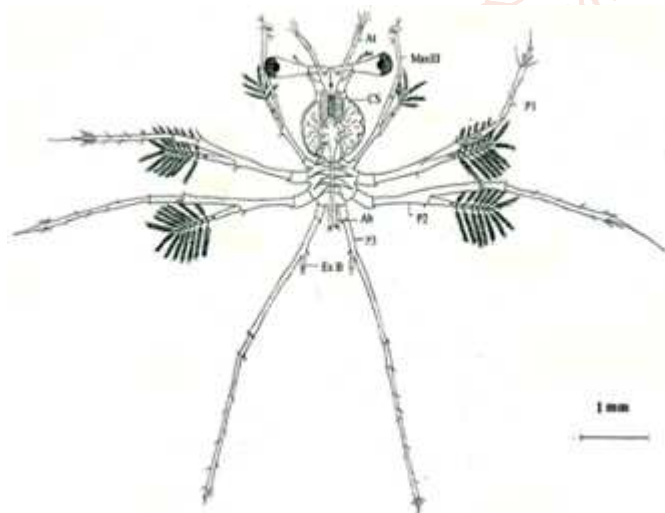


Fig.2. Phyllosoma larva after 1st moult

At – Antenna, An – Antennule, Max III – Maxillipede III, P1 – Pereiopod I, P2 – Pereiopod II, P3 – Pereiopod III, P4 – Pereiopod IV, Ab – Abdomen, CS – Cephalic shield, ExB – Exopod budding

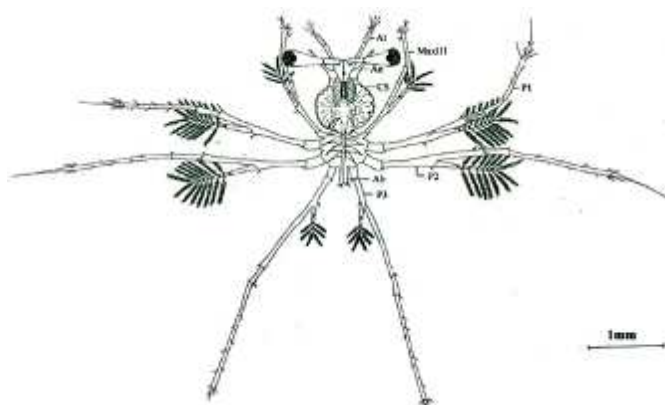


Fig.3. Phyllosoma larva after 2nd moult

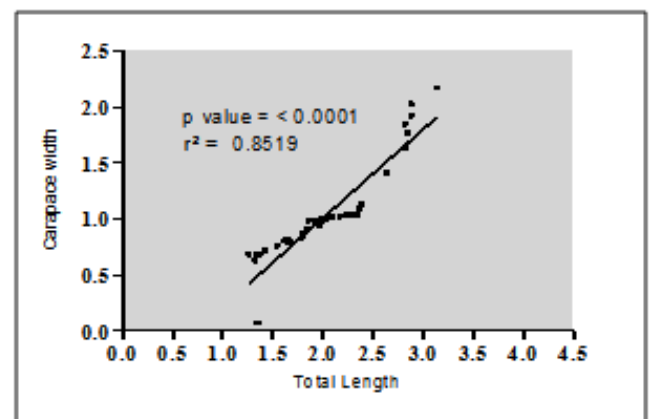


Fig.6. Percentage of growth rate of phyllosoma larvae $p < 0.0001^{***}$ (represents highly significance) done by regression analysis

Percentage of survival

There was not much mortality up to the first 10 days. For the next 10 days, the survival percentage varied between 8.14 and 5.74. After 20 days, the survival percentage started

declining from 5.39 to 2.67. From 31st day onwards, the survival percentage further declined and its was 0.01% on the 42nd day (Fig. 7).

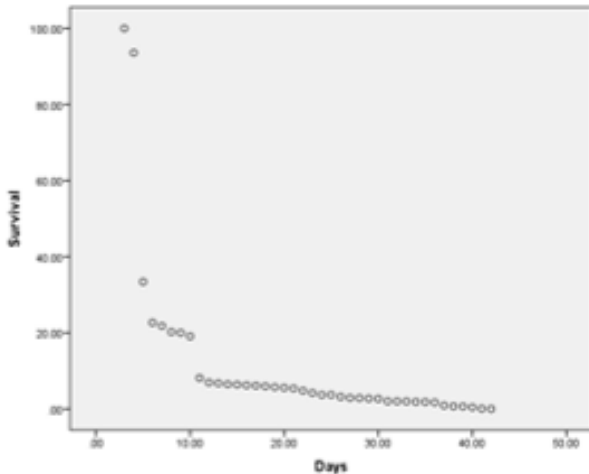


Fig.7. Percentage of survival of phyllosoma larvae

Moulting frequency

The first moult was observed after 9.00 ± 0.57 days of hatching. The second, third and fourth moultings were observed on the 19.00 ± 0.57 , 28.00 ± 0.57 and 41.33 ± 0.88 days respectively. There was significant ($p < 0.001$) difference between the days taken for each moult up to fourth (Fig.8).

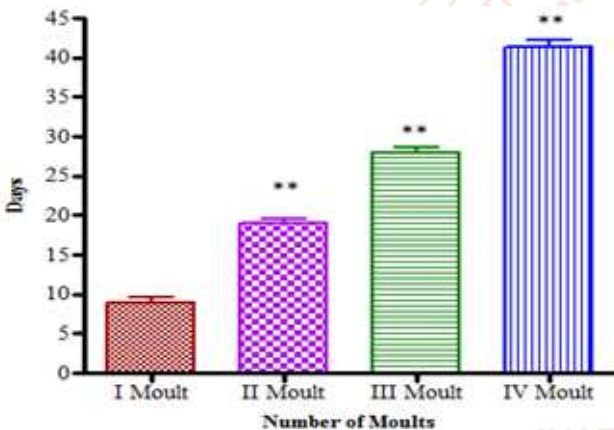


Fig.8. Moulting frequency of phyllosoma larvae

$n=3, p < 0.001^{**}$ (represents the statistical significance) done by ANOVA, followed by Turkey’s multiple comparison tests.

Fecundity

Number of larvae increased significantly ($p < 0.001$) with size of the mother lobster (Fig. 9). The spawner of 300gm size yielded $38,046 \pm 26.85$ larvae and 400 gm size brooder released $45,027 \pm 20.51$ larvae; the brooder of 550 gm size released $50, 104 \pm 57.85$ larvae.

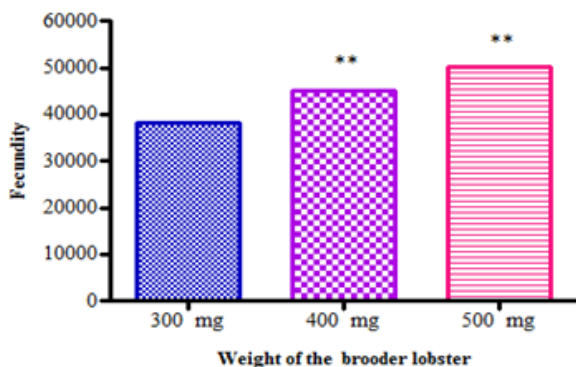


Fig.9. Relationship between brooder size and larval number

$n=3, p < 0.001^{**}$ (represents the statistical significance) done by ANOVA, followed by Turkey’s multiple comparison tests

DISCUSSION

When comparing the present result, with Sankolli and Shenoy (1973) findings, the antennule, antennae, mandible, first maxilla, second maxilla and second maxilliped had similar morphological changes. But the following parts observed by them showed variation with the present work. The first maxilliped was undeveloped, the third maxilliped was uniramous, the exopod of first and second pereopod had 8 segments and 7 pairs of plumose setae. But in the present study, the larvae exhibit developed first maxilliped and biramous third maxilliped. In the present case, the exopod of first and second pereopod had 7 segments and 6 pairs of plumose setae each. Again, the present work showed similar morphological changes in the following parts such as eye, eye stalk, antenna, mandible, first maxilla, second maxilliped, and third maxilliped. They observed four apical setae in the second maxilla, but no such setae were observed in the present study. Nine pairs of setae were recorded by them in the first and second pereopod, but in the present study only 7 pairs of setae were observed.

A study conducted by Radhakrishnan and Vijayakumaran (1993) on phyllosoma larvae of *Panulirus homarus* showed an segmented eye stalk during the first moult. The present study also confirmed their findings. The carapace width observed by them was 0.72 mm and in the present study it was 0.68 mm. The early stages of the *Panulirus echinatus* moulted eight times and morphological changes of each appendage were described Abrunhosa *et al.* (2008). Again a short span study conducted by Radhakrishnan *et al.*, (2009) on the survival and growth of phyllosoma larvae of the tropical spiny lobster, *Panulirus homarus* maintained with microalgae *Nannochloropsis salina* culture treatments and without microalgae culture, the author reported that the phyllosoma larvae moulted nine (I- VIII stages) and six times (I – V stages) in the microalgal and non-algal systems, respectively. They found the exopod of fourth pereopod appeared to become setose in the V stage, the antennule becomes four segmented in the VI stage, 5th pereopod appeared as small bud in VII stage, 5th pereopod budding becomes elongated and formed biramous in stage VIII respectively. At the end, the total length was 5.25 ± 0.04 and carapace width was 3.75 ± 0.04 but at the end of the present study after 4th moult, the total length was 2.89 ± 0.62 mm and carapace width was 2.01 ± 0.14 mm respectively and also showed the entire morphological characters of phyllosoma larvae up to 4th moult.

The spawner size also showed direct relationship with nauplii or egg production. The same fact has been observed in *P.monodon* by (Babu *et al.*, 2001b). About 66% of the breeders belonged to the size group of 61-80 mm CL and this is the dominant size group in the fishery as evident from the export of live lobsters. This size group may be contributing more to the reproduction and recruitment in *P. homarus*. This may be due to the deposition of more nutritional reserve in the body or its increased age.

The fast growth and high survival of early stage phyllosoma larvae of *P. homarus* in culture system circulated with *N. salina* and continuous enrichment of *Artemia* by feeding on *N.salina* in the micro-algal system was reported by,

Radhakrishnan *et al.* (2009). The present study was done under normal condition. The percentage of survival had been declining from 31st day onwards. It was also noticed that the major cause of the larval mortality was the lack of suitable food for different larval stages (Sarasu and George, 1993; Abrunhosa, 2008).

ACKNOWLEDGEMENT

The authors are thankful to the University Grants Commission, Government of India for providing necessary funds for operating this project and Manonmaniam Sundaranar University, Tirunelveli, TamilNadu for providing Laboratory facilities.

REFERENCES

- [1] Abrunhosa, F. A., A. P. Santiago and J. P. Abrunhosa. 2008. The early phyllosoma stages of spiny lobster *Panulirus echinatus* Smith, 1869 (Decapoda: Palinuridae) reared in the laboratory. *Braz. J. Biol.*, 68(1): 179-186.
- [2] Babu, M. M, and M. P. Marian. 1998. Live transport of gravid *Penaeus indicus* in Coconut mesocarp dust. *Aquacultural Engineering*, 18:149-155.
- [3] Babu, M. M, and M. P. Marian and M. R. Kitto. 2001a. A cradle aeration system for hatching *Artemia*. *Aquacultural Engineering*, 24: 85-88.
- [4] Babu, M. M., S. Lazarus, M. P. Marian and M. R. Kitto, 2001b. Factors determining spawning success in *Penaeus monodon*, NAGA, The ICLARM, Quarterly (Vol.24) Nos. 1& 2.
- [5] Dilip K., M. Vijaykumar, T. Senthil Murugan, J. Santhanakumar, T. S. Kumar, N. V. Vinithkumar and R. Kirubakaran. 2010. Survival and growth of early phyllosoma stages of *Panulirus homarus* under different salinity regimes. *J. Mar. Biol. Ass. India*, 52(2): 215-218.
- [6] Kittaka, J., 1988. Culture of the palinurid *Jasus lalandii* from egg to puerulus. *Bull. Japanese Soc. scient. Fish.*, 54(1):87-94
- [7] Kittaka, J. and E. I. Kegami, 1988 (cf. a). Culture of the palinurid *Palinurus elephas* from egg to puerulus. *Bull. Japanese Soc. Scient. Fish.*, 54(7):1149-1154.
- [8] Kittaka, J., M. Iwai and M. Yoshimijra, 1988 (cf. b). Culture of a hybrid of spiny lobster genus *Jasus* from egg stage to puerulus. *Bull. Scient. Japanese Soc. Fish.*, 54 (3):413-418.
- [9] Lazarus, S., S. G. P. Vincent and M. M. Babu. 2000. FAA-Rich Marine Microalgae as energy fuel for the early feeding of phyllosoma larvae of the spiny lobster, *Panulirus homarus*. In *National symposium on phycology in the new millennium*, Centre for Advanced studies in Botany, University of Madras, Tamil Nadu, India, p.30.
- [10] Maharajan, A., M. Vijayakumar, M., S. Rajalakshmi, P. Jayagopal, M. S. Subramanian and M. C. Remani. 2012. Fecundity and viability of eggs in wild breeders of spiny lobsters, *Panulirus homarus* (Linnaeus, 1758), 2012. *Panulirus versicolor* (Latrielle, 1804) and *Panulirus ornatus* (Fabricius, 1798). *J. Mar. Biol. Ass. India*, 54:201-209.
- [11] Radhakrishnan, E. V. and M. Vijayakumar. 1995. Early larval development of the spiny lobster *Panulirus homarus* (Linnaeus, 1758). *Crustaceana.*, 68 (2):151-159.
- [12] Radhakrishnan, E. V., Rekha D. Chakraborty, R. Thangaraja and C. Unnikrishnan, 2009. Effect of *Nannochloropsis salina* on the survival and growth of phyllosoma of the tropical spiny lobster, *Panulirus homarus* L. under laboratory conditions. *J. Mar. Biol. Ass. India*, 51 (1): 52 – 60.
- [13] Radhakrishnan, E. V. and M. Vijayakumar. 1993. Early larval Development of the spiny lobster *Panulirus homarus* (Linnaeus, 1758) reared in the laboratory. *Proceeding of the fourth International Workshop on Lobster Biology and Management*.
- [14] Sankolli, K. N. and Shenoy. 1973. On the laboratory hatched six phyllosoma stages of *Scyllarus sardidus* (stimpson). *Mar. Biol. Assoc. India*; 15(1): 218-226.
- [15] Sarasu, T. N. and M. J. George. 1993. Larval Biology of spiny lobsters of genus *Panulirus* *CMFRI Spl. Publ.*, 56:45-47.
- [16] Vijayakumar, M., T. Senthil Murugan, M. C. Remany, T. Mary Leema, J. Dilip Kumar, J. Santhanakumar, R. Venkatesan, M. Ravindran. 2005. Captive breeding of the spiny lobster, *Panulirus homarus*. *New Zealand Journal of Marine and Freshwater Research*, vol. 39: 325-334.